## Mt. San Antonio College Microbiology 22 Lab Schedule for Fall 2016 Mon/Weds. Split Lab Sections ONLY

Wk	Mon. Date	Mon. Lab Activities: 1 hour 25 minutes (No break)	Weds. Date	Weds. Lab Activities: 1 hour 25 minutes (No break)
1	Aug. 29	Orientation with Introductions & Safety Rules/Regulations	Aug. 31	Orientation with Pathogen Groups
2	Sept. 5	HOLIDAY: LABOR DAY!	Sept. 7	<b>Exercise #1:</b> <b>The Microscope</b> Objectives: Learn the parts of the compound microscope and their functions. Learn how to safely transport, clean, and store the microscope. Learn to observe various specimens on slides using the low power, high power, and oil immersion objectives. Learn to identify the three basic morphologies of bacteria, and some of their characteristics.
3	Sept. 12	<i>Exercise #1 continued</i> : The Microscope and Introduction to Bacteria!	Sept. 14	Quiz 1 (lab safety/regulations         & Lab Exercise 1)         Exercise #2:         Culturing the Environment and         Your Hands!         Objectives: Learn terms related         to culturing microorganisms,         demonstrate aseptic techniques and         lab procedures, describe colony         characteristics, compare bacterial         growth on your samples, and         describe why agar is used in culture         media.

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4	Sept. 19	Exercise #3:         Media, Aseptic Techniques         and Handling Cultures         Objectives: Carry out the         technique for aseptic removal         and transfer of microorganisms         for subculturing. Correctly         sterilize inoculating instruments         using the Bacti-cinerator.         Evaluate bacterial growth on         slant and broth subcultures.         Write-Up this Lab; due in 3         weeks at the beginning of lab         class!         (PG1)*	Sept. 21	Exercise #4: Simple Staining Objectives: Prepare bacterial smears for microscopic visualization. Perform simple staining from liquid and solid media. Compare shapes and arrangements of bacterial cells.
5	Sept. 26	Quiz 2 (Lab Exercises 2 & 3 + PG1) Simple Staining continued to completion.	Sept. 28	<i>Exercise #5</i> : Gram Stain Objectives: Become proficient in making and viewing Gram stain, acid- fast stain, and spore stain slides.
6	Oct. 3	<u>Exercise #5 continued:</u> Acid-fast and Spore Stain (PG 2)*	Oct. 5	Unknown staining

ate	1 hour 25 minutes		
	1 nom 20 minutes	Date	1 hour 25 minutes
	(No break)		(No break)
ct. 10	Quiz 3 (Lab Exercises 4 & 5 + PG2), Unknown staining continues to completion	Oct. 12	Fungi Lab ( <i>Exercise #6</i> ) Objectives: Learn to distinguish among types of hyphae. Learn yeast morphology (budding, pseudohyphae). Learn to distinguish among several different fungi based on their physical characteristics. Learn vocabulary associated with fungi parts. Learn the importance of HardyCHROM differential culture media. See the parts of lichen. (PG 3)*
ct. 17		Oct. 19	
	Quiz 4 (fungi lab+ PG3)	000015	Quiz 5 (protozoa lab+ PG4)
	<b>Protozoa observations</b> (Exercise #7)		Worms/Arthropods Lab observations ( <i>Exercise #8</i> )
	Objectives: Compile information about several protozoa, and learn to distinguish among them based on unique physical characteristics.		Objectives: Compile information about several worms and arthropods, and learn to distinguish among them based on unique physical characteristics.
	(PG 4)*		(PG 5)*
ct. 24	. Quiz 6 (worms/arthropods lab+ PG5) Exercise #9: Pure Culture Techniques Objectives: Isolate bacteria by using streak plate and pour plate techniques. Prepare and maintain a pure culture.	Oct. 26	Subculture for pure culture challenge points AND <u>Exercise #10:</u> Perform isolation of "unknown bacterial pathogens" on specialized media (Complete parts A, B, & C) Objectives: Learn how to obtain clinical specimens, become familiar with culture media used to isolate pathogens, and make representative cultures.
c	t. 17 	t. 17 Quiz 4 (fungi lab+ PG3) Protozoa observations (Exercise #7) Objectives: Compile information about several protozoa, and learn to distinguish among them based on unique physical characteristics. (PG 4)* t. 24 . Quiz 6 (worms/arthropods lab+ PG5) Exercise #9: Pure Culture Techniques Objectives: Isolate bacteria by using streak plate and pour plate techniques. Prepare and maintain a pure culture.	t. 17       Quiz 4 (fungi lab+ PG3)       Oct. 19         Protozoa observations (Exercise #7)       Objectives: Compile information about several protozoa, and learn to distinguish among them based on unique physical characteristics.       Oct. 26         (PG 4)*       Oct. 26         t. 24       Oct. 26         . Quiz 6 (worms/arthropods lab+ PG5)       Oct. 26         Exercise #9:       Pure Culture Techniques Objectives: Isolate bacteria by using streak plate and pour plate techniques. Prepare and maintain a pure culture.

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10	Oct. 31	<ol> <li>Record your results from previous lab</li> <li>Perform <i>Exercise #11:</i> More Lab Tests (of the isolated "unknown bacterial UTI specimen")</li> <li>Objectives: Demonstrate biochemical characteristics, motility, and oxygen requirements of microorganisms.</li> </ol>	Nov. 2	Quiz 7 (Lab Exercise 9 + PG 6)Record the results of the MoreLab Tests of your UTI mysterymicrobeANDThe API 20E System(Exercise #12) is put to use forhelping ID our "UTI pathogensObjectives: Understand the meaning of"enteric," compare the media andconventional tube methods you've used,and learn to use the API 20E system.
11	Nov. 7	Record API 20E results to key out your "Unknown pathogen" <u>Exercise #13:</u> Confirmation of the UTI results: confirm results! Objectives: Use results of exercises 10-13 to identify your mystery microbe. (PG 7)*	Nov. 9	Quiz 8 (Lab Exercises 10, 11, 12 and 13 + PG 7) Start 'Control of Microbes' with 3 Labs 1. <u>Exercise #14:</u> Antimicrobial Susceptibility Testing. Objectives: Learn to perform antimicrobial susceptibility tests using paper discs. Test effects of dyes and metals.

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12	Nov 14		Nov 16	
12	100.14	Record results of your	100.10	
		team's		Record results of the entire
		S, I, R ratings.		class's S,I,R ratings and discuss.
		AND		AND
		2. Exercise #15: Perform		Record class results of Heat,
		Heat, Cold, Drying, Radiant		Cold, Drying, Radiant Energy
		Energy Lab		Lab
		Objectives: Demonstrate the effectiveness of these control methods against common microorganisms.		(PG 8)*
13	Nov. 21		Nov. 23	
		3. <u>Exercise #16:</u> Perform		<b>Record results of antibacterial</b>
		Antibacterial Products		products tests.
		Testing (each student brings		· Ori- 0 (Lab Errorian 14, 15, 9
		a lavorite antiseptic or disinfectant to test (2)		+ $Quiz 9$ (Lab Exercises 14, 15 $\&$ 16 + PG8)
		Objectives: Compare		10 11 (0)
		effectiveness of disinfectants,		
		antiseptics, and cleaning products.		
		time, concentration, and species.		
		Write-Un this Lah Ex. 16		
		(optional); due on Dec. 7th at		
		the beginning of lab class- to		
		replace 1 <sup>st</sup> Write-Up score to		
		possible 40 pts.!		

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14	Nov. 28	<u>Exercise #17:</u> Simulation of Disease Spread: an HIV epidemic + HIV antibody detection with enzyme linked immunosorbent (ELISA) assay! Objectives: Take part in synthetic epidemic, use ELISA to detect HIV antibodies, and use epidemiology to learn source of epidemic. (PG 9)*	Nov. 30	<i>Exercise #18:</i> Transformation of <i>E. coli</i> experiment Objectives: Perform genetic transformation of <i>E. coli</i> with a jellyfish gene that codes for a fluorescent protein.
15	Dec. 5	Record Transformation results         Exercise #19: Perform         Milk/Water lab (each student brings)         Objectives: Perform bacteriological testing procedures for water and milk, including testing for coliforms, membrane filtration, and plate counts.	Dec. 7	Record milk/water results Lab Quiz 10 ( Lab Exercises 17, 18 & 19 + PG 9 )

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16		FINAL EXAMS WEEK The Pathogen Test is your Lab Finalcheck the final exam schedule for the time it will be given based on your lab meeting time during the semester.		
		YOUR LAB'S PATHOGEN TEST WILL BE DATE: TIME:		

\* Date that **pathogen group** (**PG#**) should be completed so you'll be ready for the next quiz! Bring your charts for discussion/sharing.